

Disclosures - Non

Molecular Diagnosis of CNS Infectious Diseases

**World Congress of Neurology
Santiago de Chile
CNS Infections Teaching Course**

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31st October 2015
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Objectives

- Learn about the use and abuse of molecular diagnostic testing for CNS infections
- See some specific examples
- Examine the latest developments in this area which may become routine in future

- Illustrative Interactive Cases
- Recent Research
 - Our group and others

Key Messages

- Put effort into getting the right samples
- Only asks for tests where the diagnosis is possible
- PCR is appropriate for many CNS infections
 - Viruses, bacteria, TBM
 - HIV disease
 - But not all infections
- Think about which other tests may be helpful
- Future
 - Role of dual infections
 - Deep sequencing approaches may help identify new pathogens
 - Host response approaches will also prove useful

Learning Points from this case

- Encephalitis Presentation can be subtle, fluctuant
- Clues
 - Lethargy Drowsiness, Sleeping 15hrs/day, Severe Headache, Hiccoughs, SIADH
- Beware “atypical pneumonia”
- Molecular diagnostic tests don’t work unless you DO the investigation (LP), chase the result and act on it

Why encephalitis is missed

- Wrongly attributing a patient's **fever and confusion**
 - “urinary tract infection”
 - “chest infection”
- Failure to **recognise a febrile illness**,
 - “afebrile on admission”
- **Ignoring a relative** says patient behaviour, “not quite right”
 - “Glasgow coma score =15”
- Wrongly attributing clouding of consciousness
 - “**drugs or alcohol**”
- Failure to properly investigate a patient with a fever and **seizure**
- Failure to do a lumbar puncture, even though there are no contraindications

Diagnosis of HSV in Adults

- 3 of 93 had negative initial PCR
- Approximately 3-5% of adults with HSV encephalitis will have negative initial CSF by PCR

Raschilas F, Wolff M, Delatour F, et al. Outcome of and prognostic factors for herpes simplex encephalitis in adult patients: results of a multicenter study. *Clin Infect Dis* 2002;35:254–60.

If clinical history & imaging/LP consistent with HSV encephalitis, treat even if 1st CSF negative, repeat at CSF 24-48 hours

How good is PCR for diagnosis in Children

- 38 children with HSV encephalitis (CSF PCR +ve)
- In 8 children CSF WCC <10/mm³, & Protein Normal
 - 4 of 19 samples drawn on day 1 of illness had no cells
- Initial CSF was HSV PCR negative in 8 of 33 children, before day 3 of illness

Limits of Early Diagnosis of Herpes Simplex Encephalitis in Children: A Retrospective Study of 38 Cases

Xavier De Tiège,^{1*} Bénédicte Héron,¹ Pierre Lebon,² Gérard Ponsot,¹ and Flore Rozenberg²

¹Service de Neuropédiatrie and ²Laboratoire de Virologie, Hôpital Saint Vincent de Paul, and Faculté de Médecine Cochin, Paris, France

Clinical Infectious Diseases 2003;36:1335–9

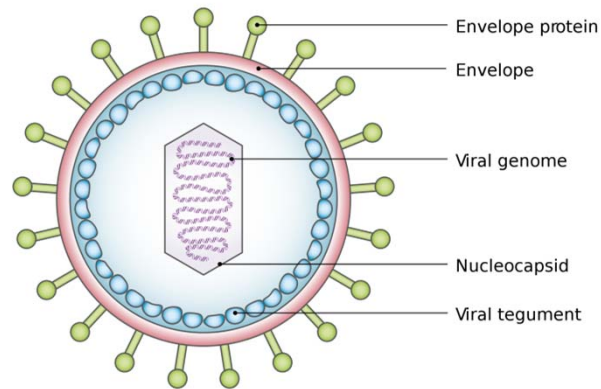
Early CSF may be normal & be PCR Neg
If suspicion high, continue aciclovir, repeat CSF after 24-48 hrs. Early CSF occasionally completely normal (cells protein)

How good is HSV PCR at diagnosing HSV encephalitis?

- 3 pfu or 200gEq/ml
- Sensitivity 96% and the specificity 99% when CSF is studied between 48 h and 10 days from the onset of symptoms
 - (Lakeman and Whitley, 1995; Tebas et al., 1998).

Diagnosis of CNS Infections – the basics

Pathogen



Host



Composition

Whole virion

Nucleic Acid

- DNA
- RNA

Antigens / proteins

Detection through:

Culture / Electron Microscopy

PCR

(Next generation sequencing)

ELISA / Immunofluorescence

Host Response

Antibodies by ELISA
(mRNA expression)

How to ensure you get the right sample (CSF) and the right test requested

- A simple lumbar puncture pack helps
- A CNS infection “set” on the computer helps

Viral causes of encephalitis

Groups

Viruses

Herpes viruses (family Herpesviridae)

Herpes simplex virus type 1
Herpes simplex virus type 2
Varicella zoster virus type 1
Epstein-Barr virus
Cytomegalovirus
Human herpes virus 6 & 7

Enteroviruses (Family Picornaviridae)

Enterovirus 70
Enterovirus 71
Poliovirus
Coxsackieviruses, Echoviruses, Parechovirus

Paramyxoviruses (family Paramyxoviridae)

Measles virus
Mumps virus

Others (rarer causes)

Influenza viruses, Adenovirus, Parvovirus B19, Lymphocytic choriomeningitis virus, Rubella virus,

Zoonotic viruses

Rabies, other lyssaviruses
Nipah virus

Arboviruses (most are also zoonotic)

Flaviviruses (family Flaviviridae)

West Nile virus
Japanese encephalitis virus
Tick-borne encephalitis
Dengue

Alphaviruses (Family Togaviridae)

Western, Eastern and Venezuelan Equine encephalitis virus
Chikungunya

Bunyaviruses

Lacrosse virus

Coltivirus

Colorado tick fever

Vesiculoviruses

Chandipura virus

CNS infections

Bacteria

Small bacteria (mostly intracellular)

Mycoplasma
Chlamydia
Rickettsia (including scrub typhus, rocky mountain spotted fever)

Erlichiosis
Coxiella (Q fever)
Bartonella (Cat Scratch fever)
Tropherema whipplei (Whipple's disease)
Brucellosis
Typhoid fever

Spirochetes

Syphilis
Lyme Neuroborreliosis
Leptospirosis
Borrelia recurrentis (Relapsing fever)

Other bacteria

Subacute bacterial endocarditis
Listeria
Nocardia
Actinomycosis
Parameningeal infection
Abscess

Parasites

Malaria
Trypanosomiasis
Amoebic encephalitis - *Naegleria fowleri*
Cysticercosis
Echinococcus
Trichinosis
Amoebiasis

Fungi

Cryptococcus
Candidiasis
Coccidiomycosis
Histoplasmosis
North American blastomycosis

Para/Post infectious causes

Acute disseminated encephalomyelitis
Viral illnesses with febrile convulsions
Shigella
Viral infections associated with swollen fontanelle
Guillain-Barré syndrome

Non infectious diseases

Vascular

Vasculitis
Systemic lupus erythematosus
Behcet's disease
Sub-arachnoid & sub-dural haemorrhage
Ischaemic cerebrovascular accidents

Neoplastic

Primary brain tumour
Metastases
Paraneoplastic limbic encephalitis

Metabolic encephalopathy

Hepatic encephalopathy
Renal encephalopathy
Hypoglycaemia
Reye's syndrome
Toxic encephalopathy (alcohol, drugs)
Hashimoto's disease

Other

Drug reactions
Epilepsy
Hysteria
Voltage gated K channel limbic encephalitis

Non-Viral causes of encephalitis and encephalopathy

Eg Epidemiology of viral CNS infections

- Some pathogens constant across globe,
 - e.g. HSV
- Others varies with geography
 - Esp Arthropod-borne & Zoonotic
- HIV and other immunocompromise has changed epidemiology
 - CMV, EBV, Toxoplasma more important
- Vaccination has changed epidemiology in some places
 - Polio, Measles, Mumps, Japanese encephalitis

Arboviruses – growing in importance

- West Nile virus in USA and Europe
- Tick-borne encephalitis virus in Europe
- Dengue and Chikungunya spreading

- Impact of
 - climate change?
 - Changing agricultural practice
 - People movement
 - etc

Why PCR so useful in CNS Infections?

- Specific
- Very helpful in viral CNS infections (culture hard)
- Unlike blood, CSF doesn't contain PCR inhibitors (heme, endonucleases, and exonucleases), which can lead to false negatives
- CSF is sterile (unlike resp or GI tract), so detection usually means causation, so fewer false positives

PCR of the CSF especially useful for

- Some Viral CNS Infections
 - Herpes viruses (HSV, VZV, CMV, EBV, HHV6 & 7)
 - Some Enteroviruses
- Some Bacteria
 - MTB
- Some spirochetes
 - Borrelia

Limitations

- False negatives can occur
 - Pathogen no longer detectable because cleared
 - PCR inhibitors in bloody tap
 - Poor handling of sample – too many freeze thaws
- False positives can occur
 - Contamination at collection or in the lab
 - Detection of latent, non causal pathogen (e.g. EBV)

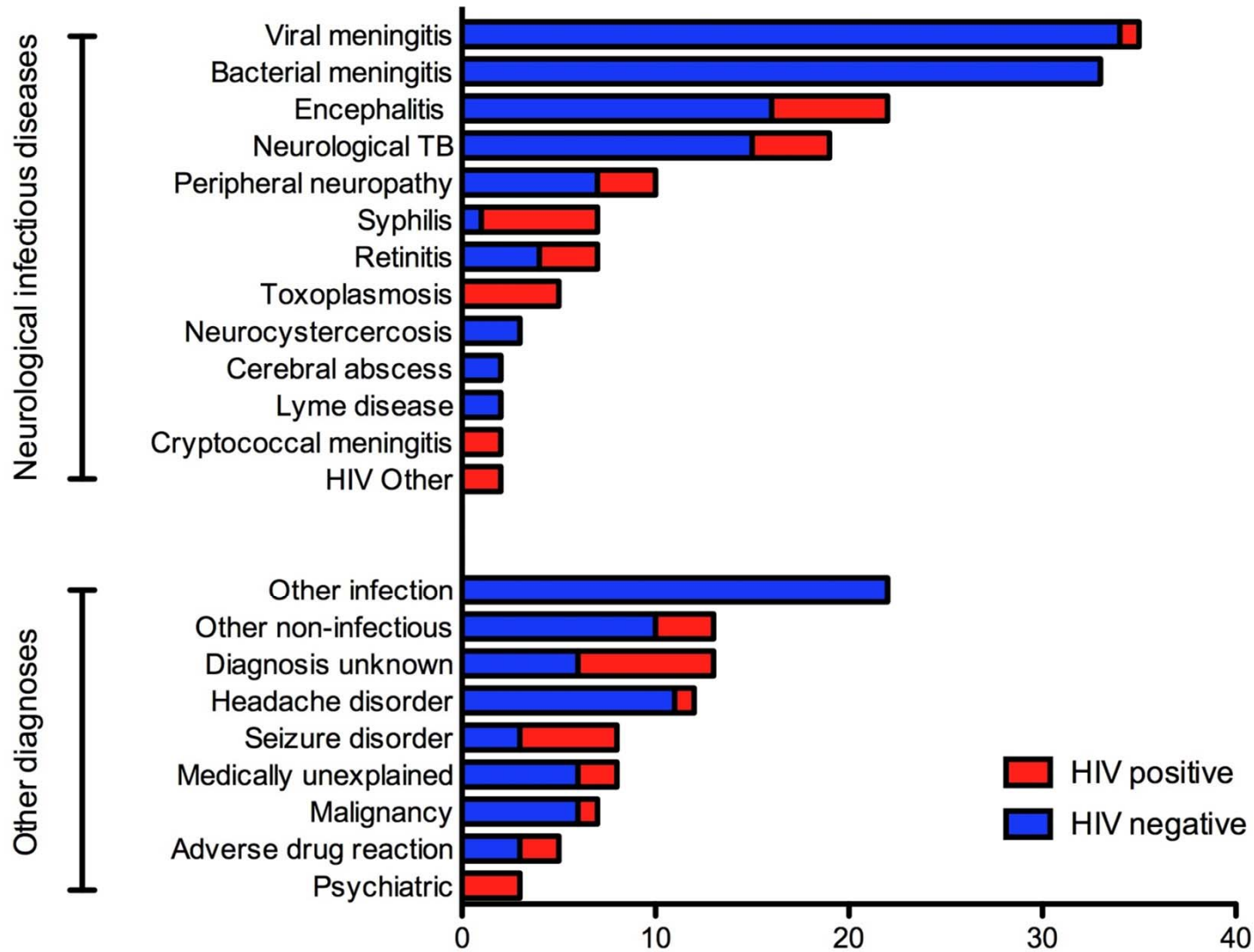
False Positives

- Only request PCR if the pre-probability testing is high
- If 5% false positive rate; then 1 in 20 tests will be falsely positive.
- If you don't think disease X is likely, what will you do if the PCR is positive for disease X!

	Toxoplasma	PCNSL	TB	PML	HAD
Clinical	Acute febrile; focal signs; Sz	Subacute dementia ; focal signs (Sz)	Subacute meningitis; focal signs, Sz	Gradual onset, visual, other focal	Chronic cognitive, motor, behaviour
Imaging	Multiple SOLs in basal ganglia and grey/white matter	Single-few SOLs periventricular / anywhere	Single-few small SOLs, or abscesses	Subcortical white matter, esp posterior; T1 hypodense	Diffuse white matter, T1 isodense
Oedema	Marked	Moderate	Marked	None	None
Enhancement	Ring	Homogeneous	Ring, and meningeal	None	None

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Imaging	<u>Multiple</u> <u>SOLs</u> in <u>basal</u> <u>ganglia</u> and grey/white matter	Single- few SOLs periventricular / anywhere	Single-few small SOLs, or abscesses	Subcortical white matter, esp posterior; T1 hypodense	Diffuse white matter, T1 isodense
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Enhancement	<u>Ring</u>	Homogeneous	<u>Ring</u> , and meningeal	None	None

Final diagnoses made in 241 patients suspected of having neurological infectious diseases



The role of a specialist neurological infectious diseases service at a secondary and tertiary hospital in North West United Kingdom Turtle L, Whiteley J, Jung A, Crocker D, Nicholson A, Beadsworth MB, Miller A, Beeching N, Solomon T, 2015

Next generation sequencing

- Limitation: PCR is specific, will only test for what you ask for
- Next generation (Deep, High throughput) sequencing is non-specific
 - Amplifies everything in the sample
 - Align all sequences
 - Ignore sequences not of interest
 - See what is left

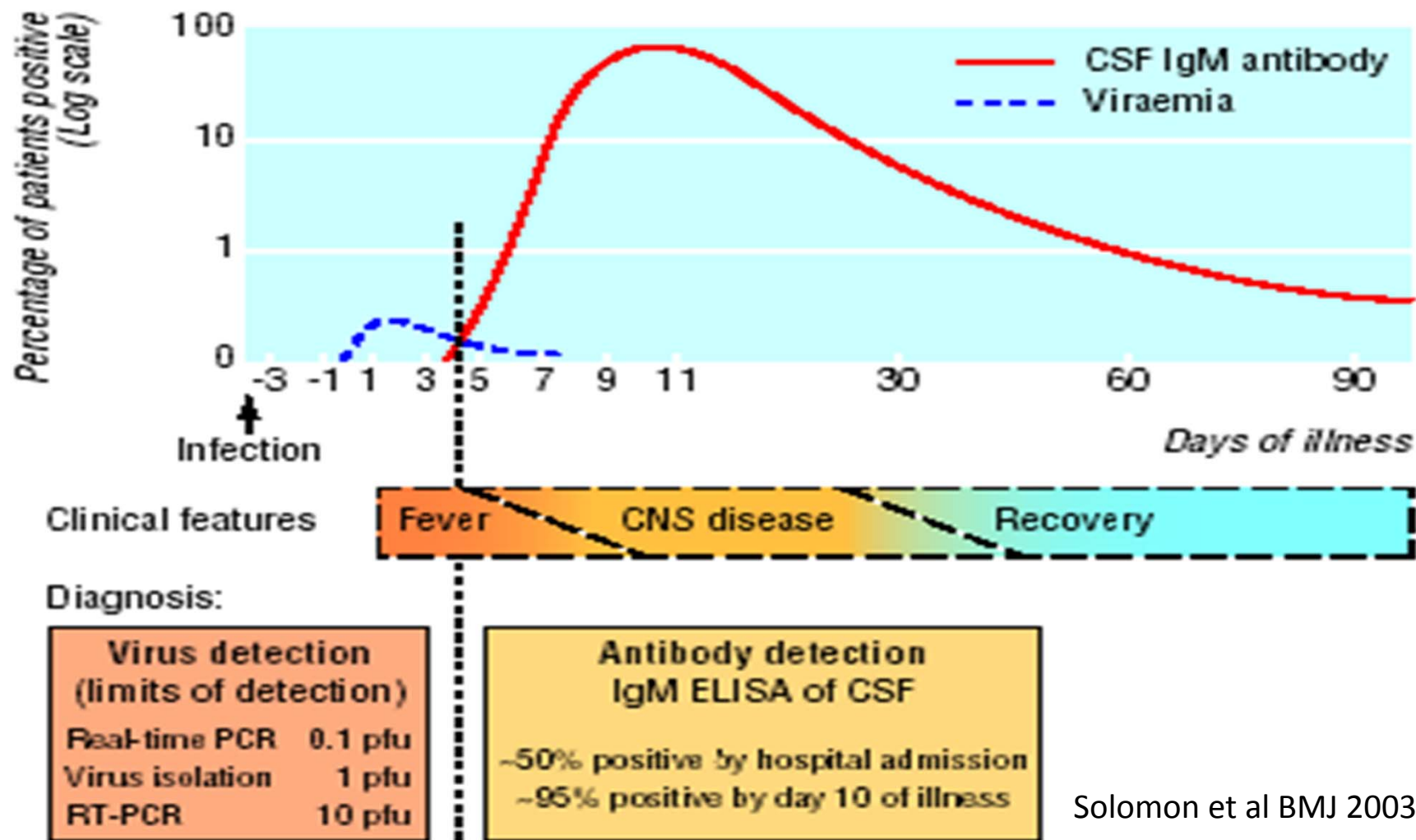
Viral Central Nervous System Infections in Children from a Malaria-Endemic Area of Malawi: a Prospective Cohort Study

- 513 children with suspected CNS infection
 - Excluded bacterial
 - 94 (18%) died.
- 163 (32%) had *P. falciparum* parasitaemia, of whom 34 died;
- 133 (26%) had at least one virus detected in the central nervous system (CNS) by polymerase chain reaction (PCR), with 43 deaths.
- Twelve different viruses were detected,
 - adenovirus most common (42 patients).
- **45 (9%) children had both parasitaemia and viral infection;**
 - 27 (35%) of 78 diagnosed clinically with cerebral malaria.



Diagnostic arbovirology

Japanese encephalitis, West Nile virus



Solomon et al BMJ 2003

Detection of NS1 antigen

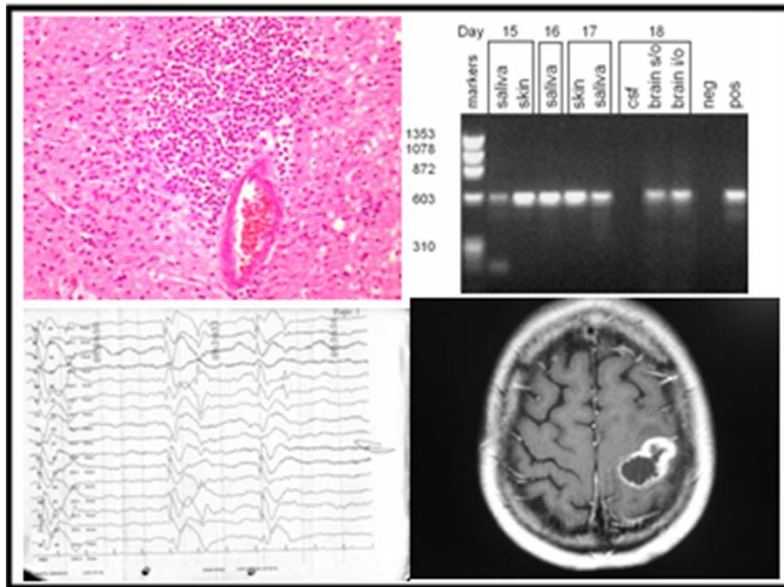
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Liverpool Neurological Infectious Diseases Course

Liverpool Medical Institution



Feedback from previous courses:

“Would unreservedly recommend to others”

“An excellent 2 days!! The best course for a long time”

Convenors: Prof Tom Solomon, Enitan Carrol, Rachel Kneen & Dr Nick Beeching

www.liv.ac.uk/neuroidcourse email braininfections@liv.ac.uk

References

- 1. Solomon T. Control of Japanese encephalitis--within our grasp? *New England Journal of Medicine*. 2006 Aug 31;355(9):869-71.
- 2. Winter PM, Dung NM, Loan HT, Kneen R, Wills B, Thu le T, House D, White NJ, Farrar JJ, Hart CA, Solomon T. Proinflammatory cytokines and chemokines in humans with Japanese encephalitis. *Journal of Infectious Diseases*. 2004 Nov 1;190(9):1618-26.
- 3. Khan SS, Smith MS, Reda D, Suffredini AF, McCoy JP, Jr. Multiplex bead array assays for detection of soluble cytokines: comparisons of sensitivity and quantitative values among kits from multiple manufacturers. *Cytometry*. 2004 Sep;61B(1):35-9.
- 4. Gould EA, Solomon T. Pathogenic flaviviruses. *Lancet*. 2008 Feb 9;371(9611):500-9.
- 5. Griffiths MJ, Shafi MJ, Popper SJ, Hemingway CA, Kortok MM, Wathen A, Rockett KA, Mott R, Levin M, Newton CR, Marsh K, Relman DA, Kwiatkowski DP. Genomewide analysis of the host response to malaria in Kenyan children. *J Infect Dis*. 2005 May 15;191(10):1599-611.
- 6. Turtle L1,2,3, Jung A4, Beeching NJ5,6,7, Cocker D8,9, Davies GR10,11, Nicolson A12, Beadsworth MB13,14, Miller AR15, Solomon T16,17,18. An integrated model of care for neurological infections: the first six years of referrals to a specialist service at a university teaching hospital in Northwest England. *BMC Infect Dis*. 2015 Sep 24;15:387. doi: 10.1186/s12879-015-1109-3.
- 7. Solomon T, Michael BD, Smith PE, Sanderson F, Davies NW, Hart IJ, Holland M, Easton A, Buckley C, Kneen R, Beeching NJ, National Encephalitis Guidelines D, Stakeholder G. Management of suspected viral encephalitis in adults--Association of British Neurologists and British Infection Association National Guidelines. *The Journal of infection*. 2012;64:347-73.
- 8. Solomon T. Encephalitis, and infectious encephalopathies. In: Donaghy M, editor. *Brain's Diseases of the Nervous System*. 12th ed. Oxford: Oxford University Press; 2009. p. 1355-428.