

XXI World Congress of Neurology  
Vienna, Austria



Teaching Course: Neurotrauma  
Challenges to nerve regeneration in  
humans:  
The Long Term Denervated Stump

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# Disclosure

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None relevant to this presentation

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# Learning Objectives

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To understand cellular mechanisms of poor recovery with proximal nerve injuries in humans

To compare and contrast regenerative therapy strategies in experimental models versus human nerve regeneration

# Challenges in PNS Regeneration

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## Intrinsic determinants of axon regeneration

- Slow rate of axonal elongation during regeneration

## Extrinsic determinants of axon regeneration

### Chronic denervation

- Changes in the pathway (i.e.. Schwann cells)

- Changes in the target (muscle or skin)

## Common issues

### Specificity of target reinnervation

- Determined through a combination of intrinsic neuronal characteristics, extrinsic pathway properties and finally target

# Acute events after injury: CNS vs PNS

## Intrinsic Factors

### CNS

#### NEURONS

Direct damage to neurons at injury site (death)

Axotomy causes neuronal death depending on proximity to cell body

#### AXONS

CNS axons fail to form growth cone and form dystrophic end bulbs (microtubule depolymerization)

~1/3 of axons sprout for ~1mm

Distal end undergoes inefficient Wallerian degeneration (myelin debris)

### PNS

#### NEURONS

Very little neuronal death

Axotomy causes neuronal death only when it is very close to cell body

#### AXONS

PNS axons form growth cone within hours (microtubules retain integrity and can bundle)

PNS axons start to regenerate shortly after injury

Retrograde injury signal

Distal end undergoes efficient Wallerian degeneration (myelin debris)

# Acute events after injury: CNS vs PNS

## Extrinsic Factors

### CNS

#### *CELLULAR*

Quick invasion of epicenter by fibroblast, vascular endothelial cells, and macrophages

Surviving host cells (astrocytes, OPC, and microglia) surround the epicenter and form a glial scar

#### *ECM*

CSPG (NG2, neurocan), fibronectin, laminin

Myelin derived inhibitors (MAG, Nogo-A, OMgp)

Guidance molecules (netrin, semaphorin, ephrin and slit families); expression changes after injury

### PNS

#### *CELLULAR*

Activation of Schwann cells and intrinsic macrophages

Invasion of blood-borne macrophages

Proliferation of blood vessels

#### *ECM*

Basal lamina is supportive of regeneration (laminin, COL4)

Myelin inhibitors are cleared

# Wallerian Degeneration

Accumulation of intra-axonal organelles

Granular disintegration of cytoskeleton

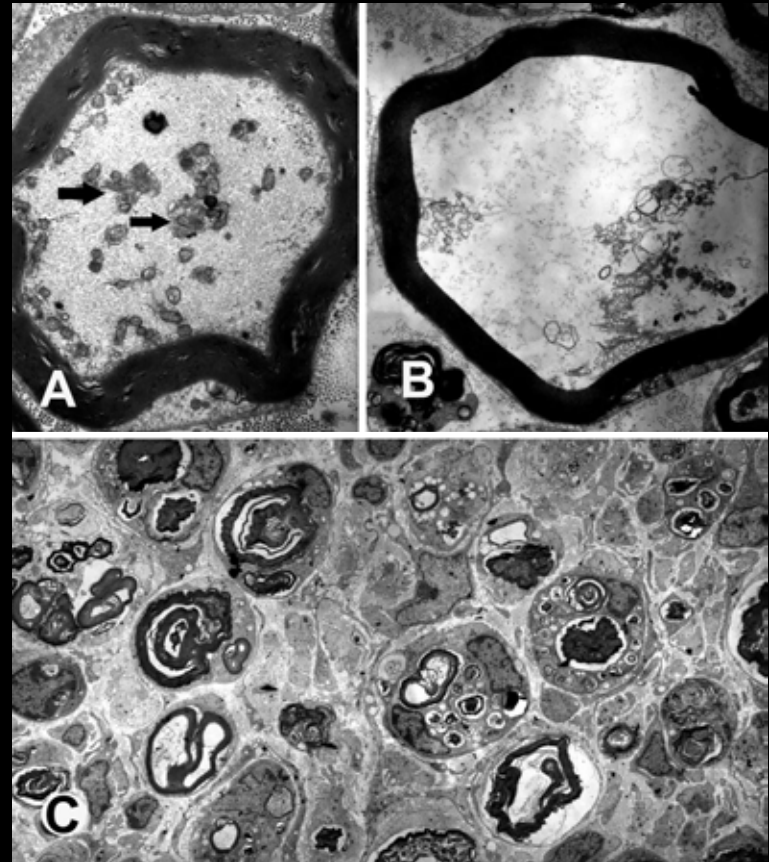
BNB/BBB breakdown

Glial changes (SC, Oligos)

Proliferation vs apoptosis

Recruitment of macrophages

Clearance of myelin ovoids



# Wallerian Degeneration

Accumulation of intra-axonal organelles

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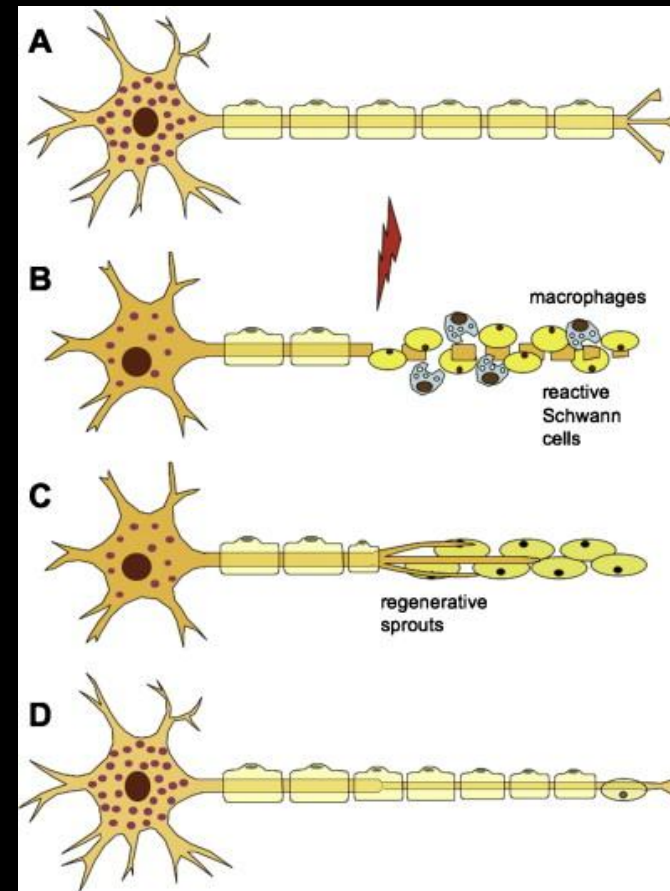
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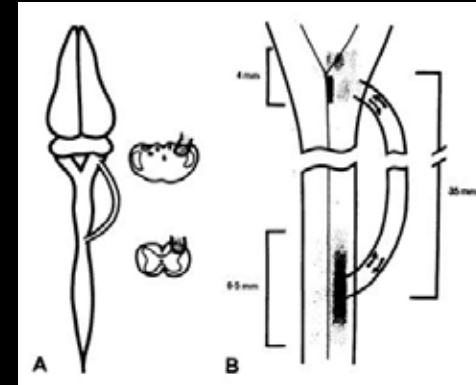
# Axonal regeneration in CNS vs PNS

Unlike CNS, axons in the PNS can regenerate

Failure to regenerate in the CNS is NOT due to an innate inability of CNS axons to regenerate

PNS environment supports regeneration

(Aguayo's experiments)



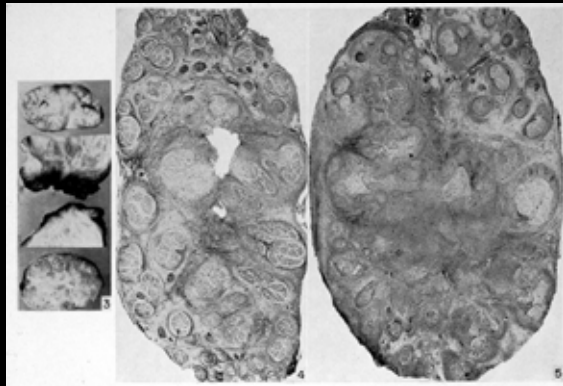
However, these observations do not correlate with clinical experience in humans

Clinical recovery after nerve trauma is often suboptimal

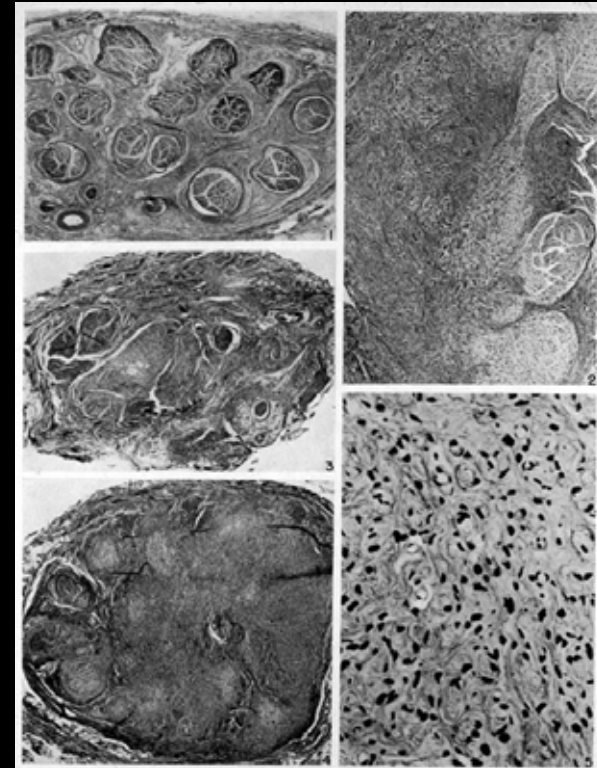
This is primarily due to “chronic denervation” (glial milieu)

# Regeneration after nerve trauma

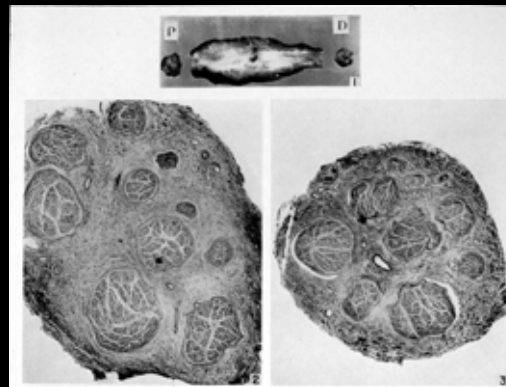
Sciatic nerve injury: no repair



Laceration to median nerve:  
Delayed repair at 10 mo



Ulnar nerve injury:  
Immediate repair at day 2



*Lyons & Woodhall 1949*

# Chronically denervated nerves do not support regeneration

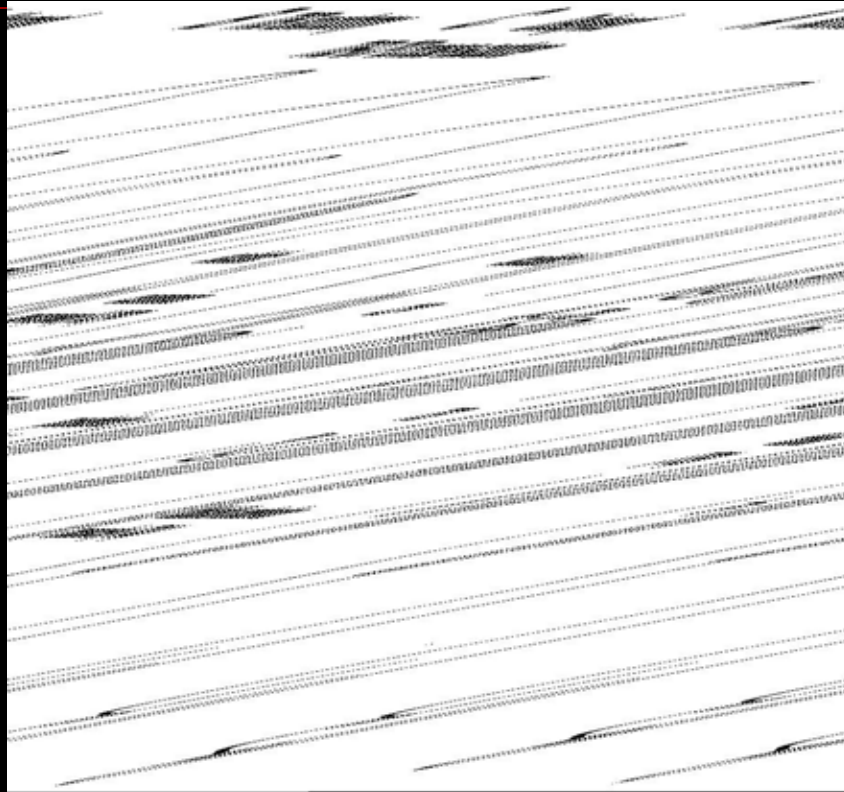
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Is this a neuron/axon problem?

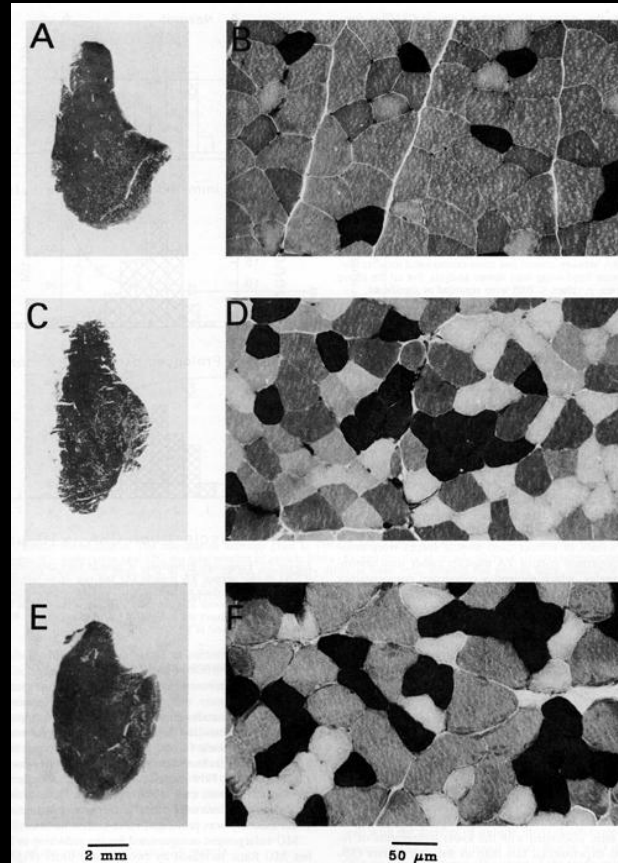
or

Is there a problem with the Schwann cells?

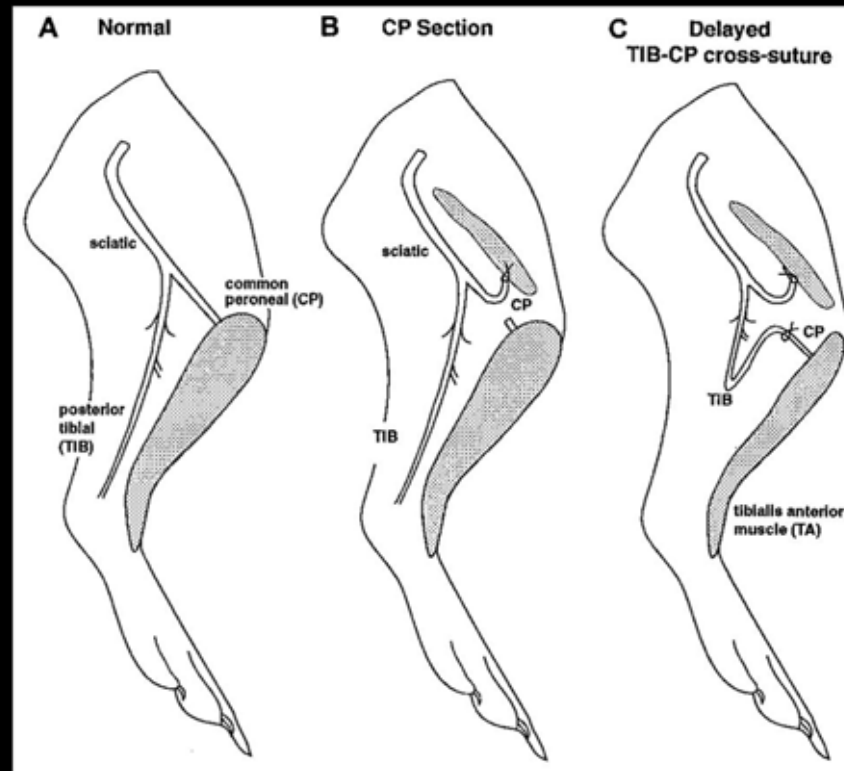
# Effects of Prolonged Axotomy



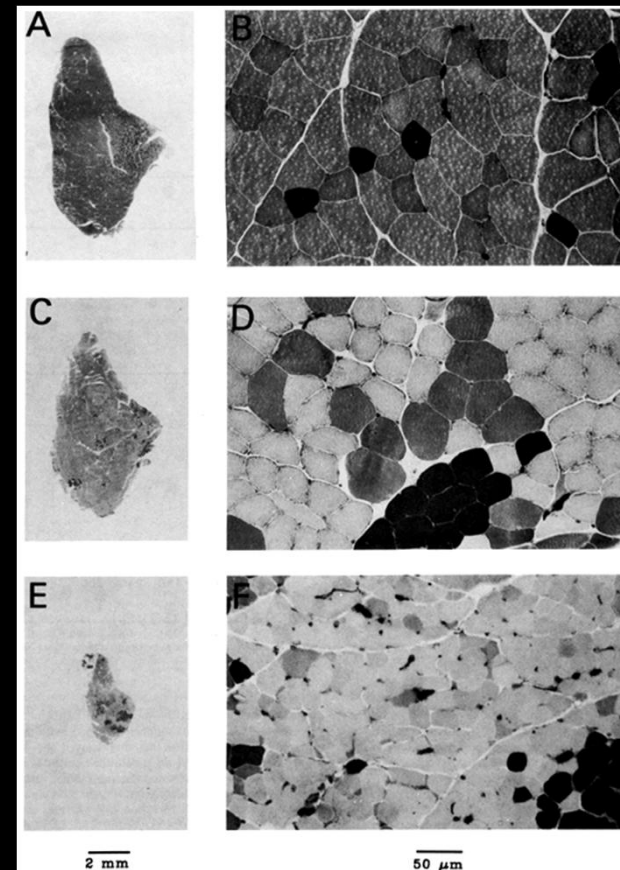
*Fu & Gordon 1995*



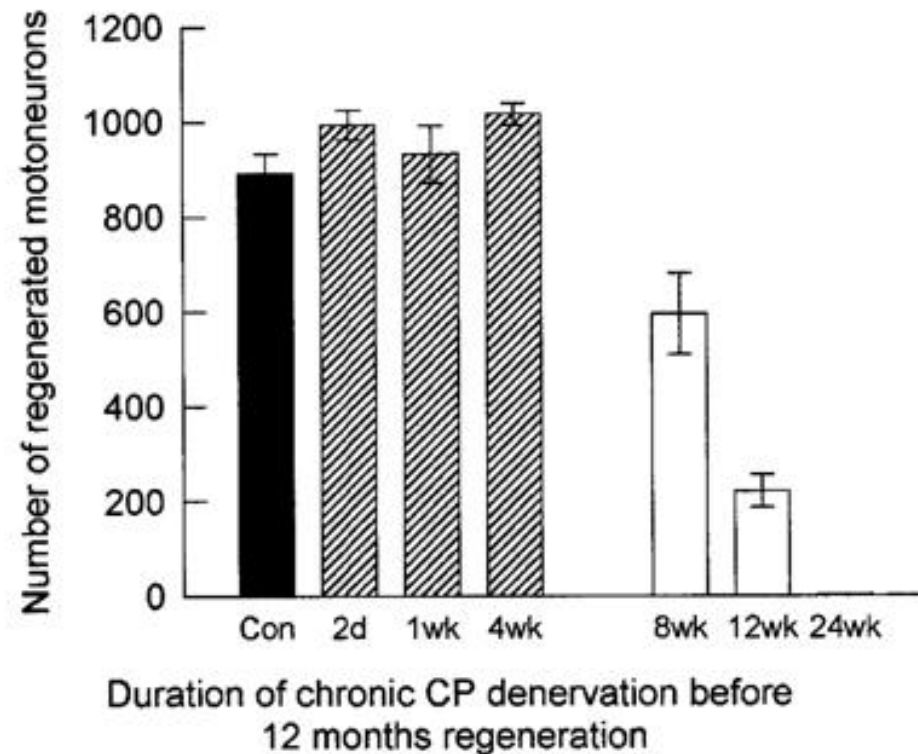
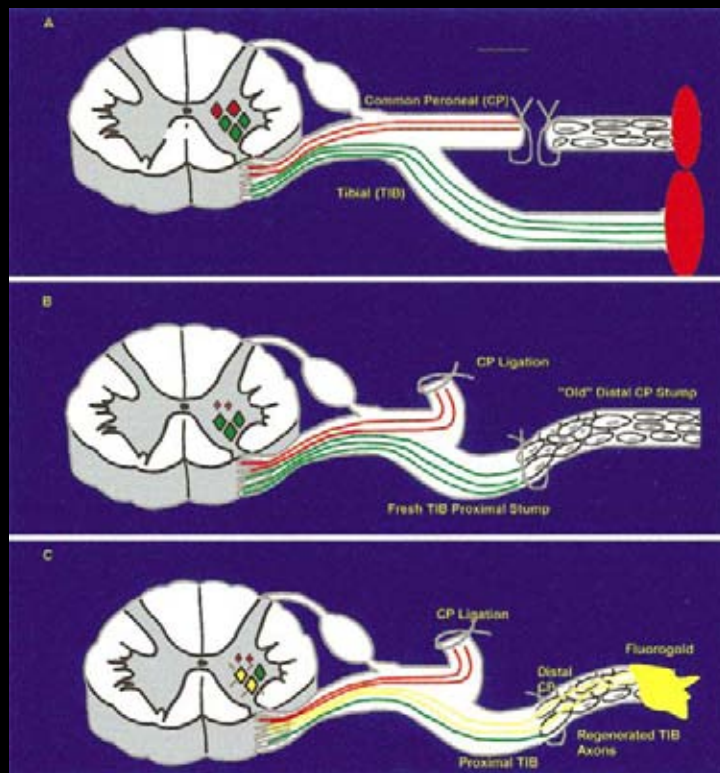
# Effects of Prolonged Denervation



*Fu & Gordon 1995*



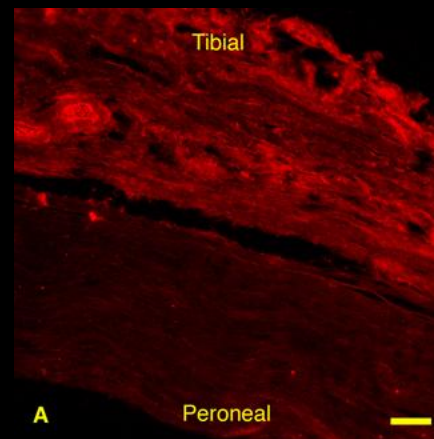
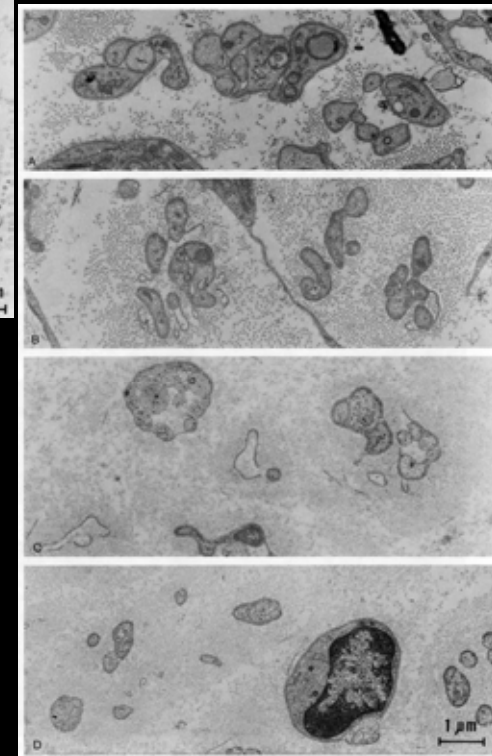
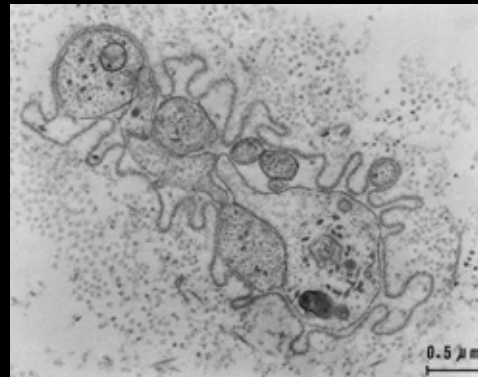
# Impaired motor regeneration following chronic denervation



*Sulaiman & Gordon 1997*

# Chronically denervated SCs lose their ability to support regeneration of axons

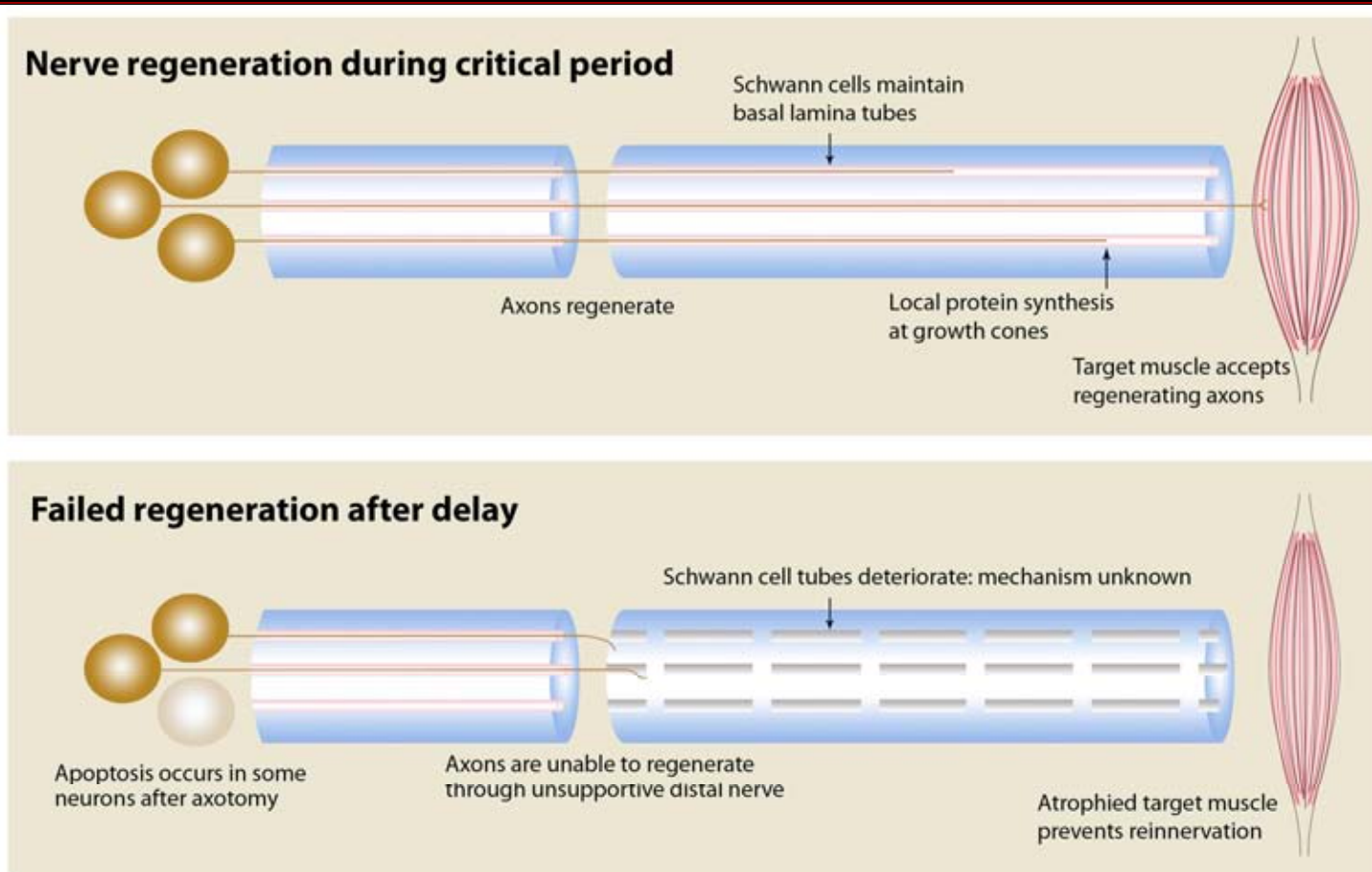
Atrophy of SCs and disappearance of Bands of Büngner  
Decline in expression of growth associated molecules (p75, erbB3) (Hall and Gordon labs) and neurotrophic factors (GDNF) (Hoke lab)  
Increased expression of CSPGs (Muir and Hoke labs)



Heine et al 2003

Peripheral Neuropathy Dyck ed 2nd ed

# Chronically denervated SCs lose their ability to support regeneration of axons

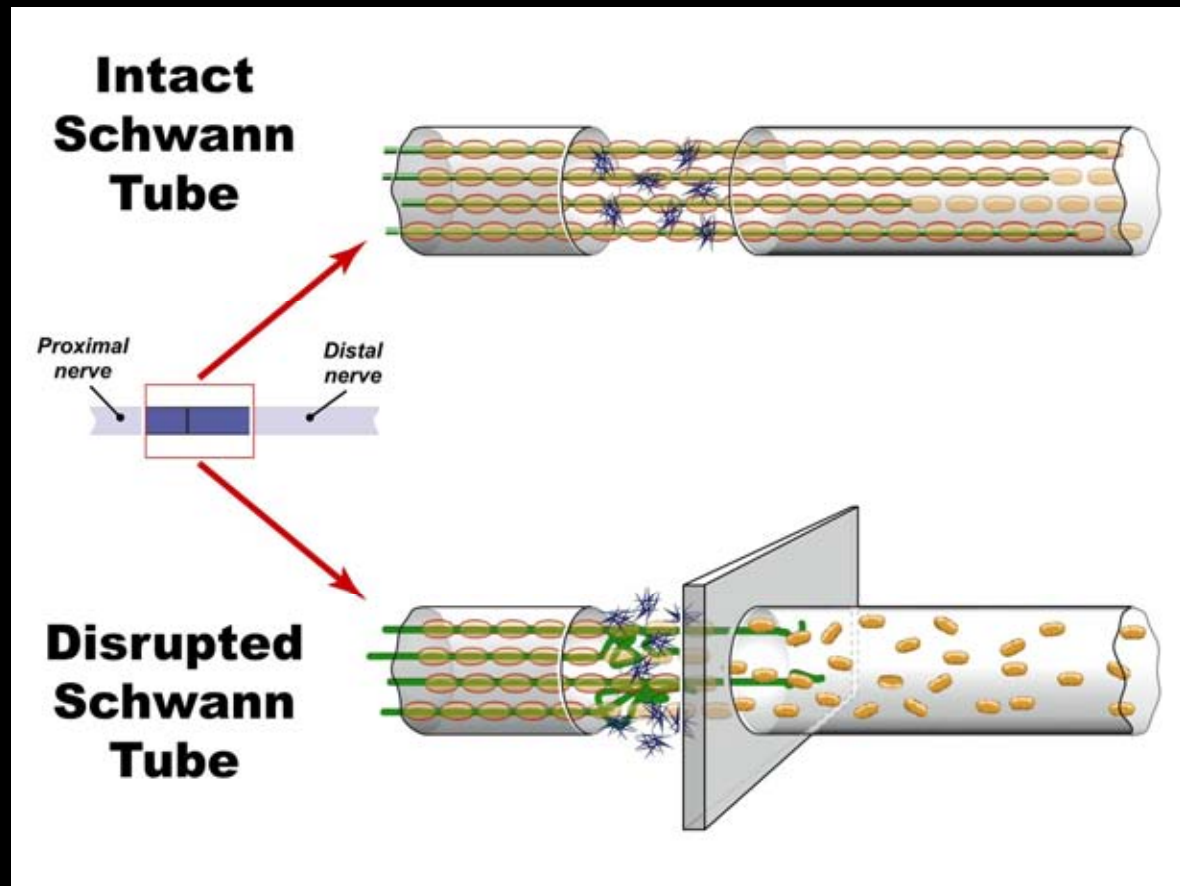




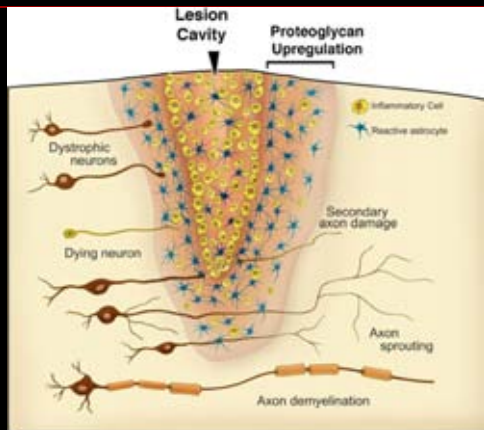
# Chronically denervated SCs lose their ability to support regeneration of axons

Acute denervation and regeneration

Chronic denervation and regeneration

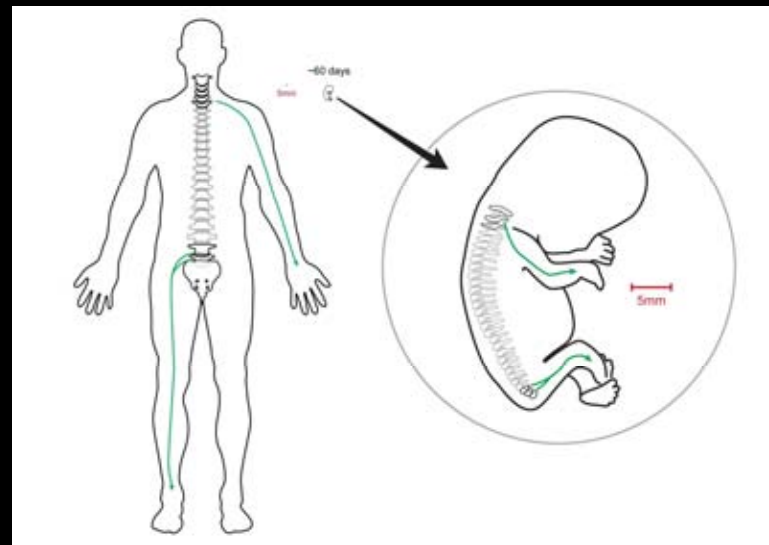


# Regenerative challenges in CNS and PNS



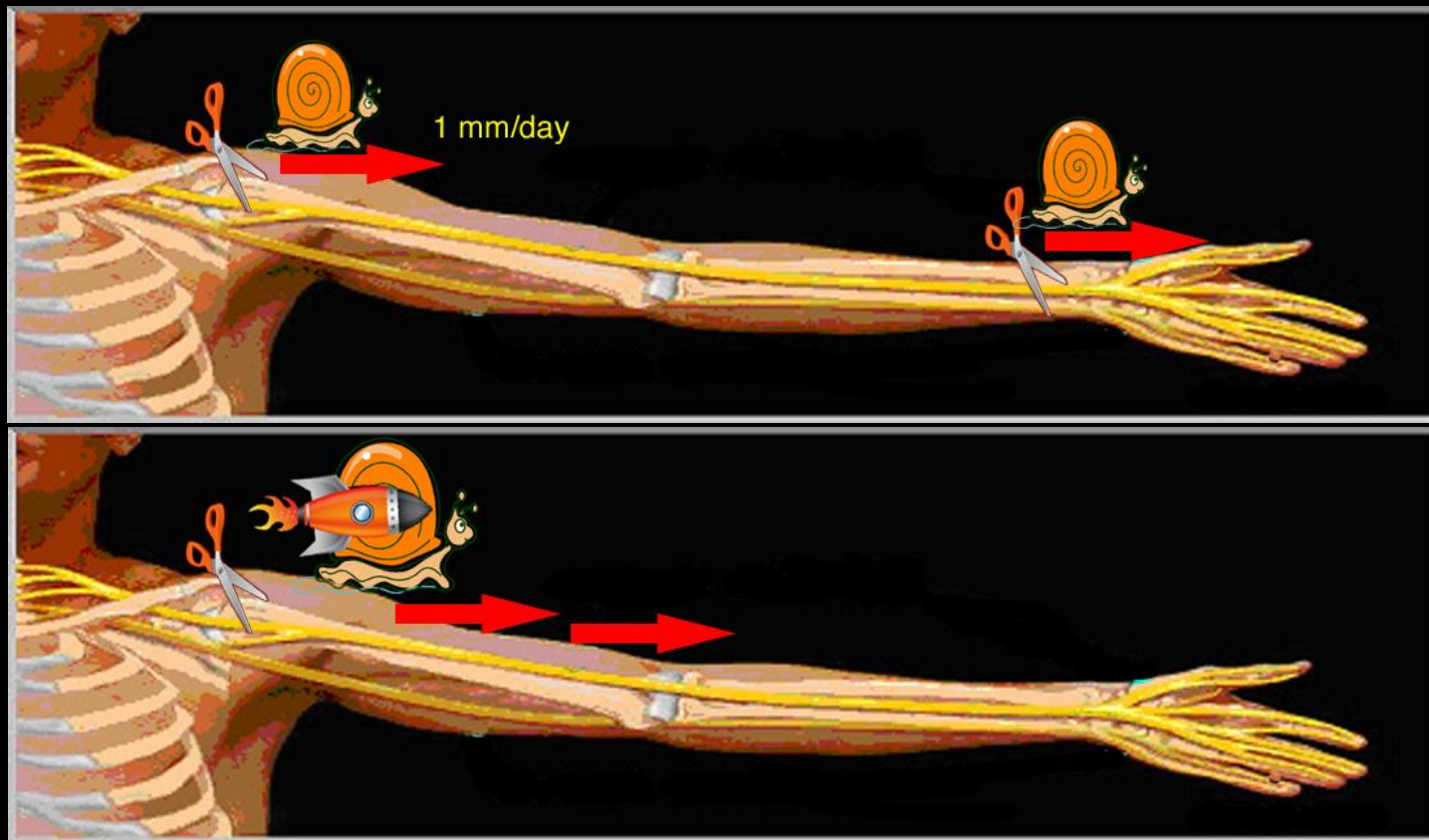
Regeneration is not the same as developmental growth:

Distance to regenerate – embryo vs adult



Glial scar  
Neuronal death  
Demyelination of spared axons

# Impact of slow rate of axonal elongation on nerve regeneration in humans



# Can we alter the rate of axonal elongation? - PNS

## Conditioning lesion

Impact on peripheral regeneration

Impact on central regeneration

## Electrical stimulation (Gordon & Brushart)

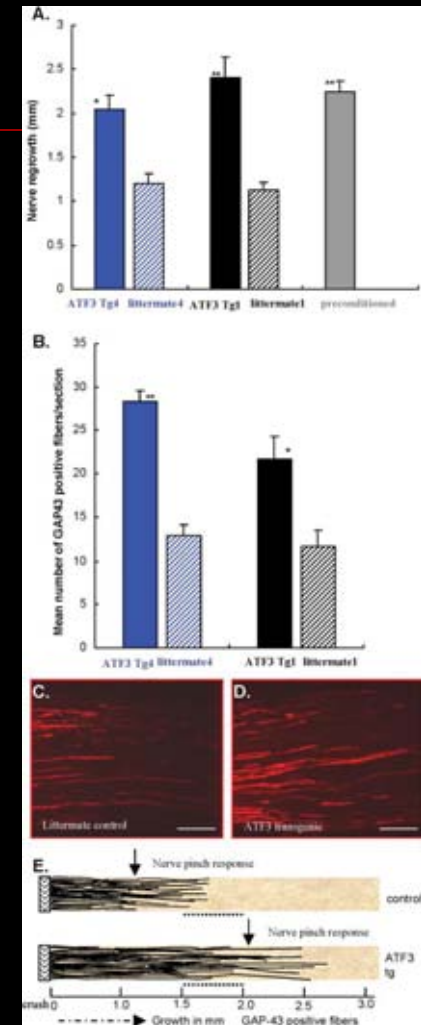
Provide pathway specificity without enhancing rate of axonal elongation

## ATF-3 over expression (Woolf)

Effect on peripheral but not central regeneration

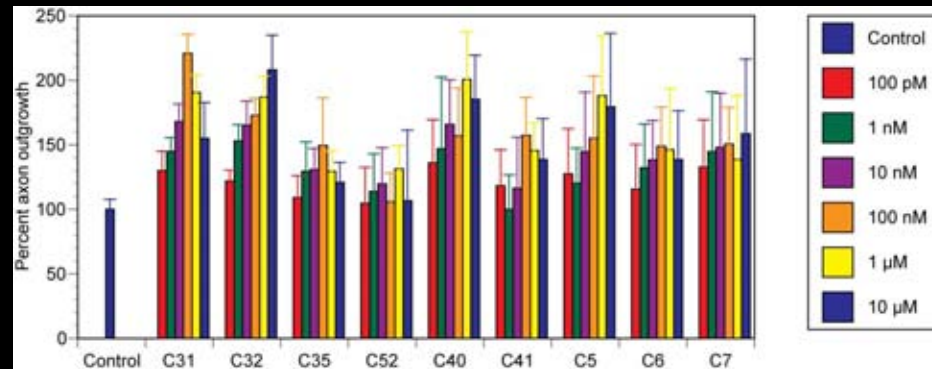
## Hsp27 over expression (Woolf)

Effect on sensory and motor regeneration

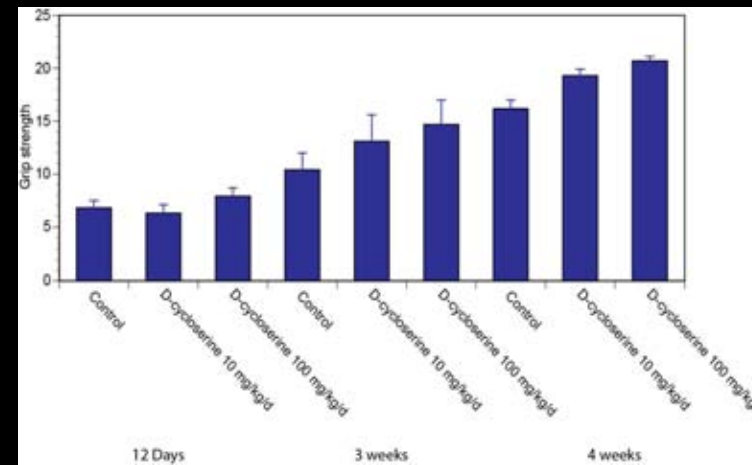
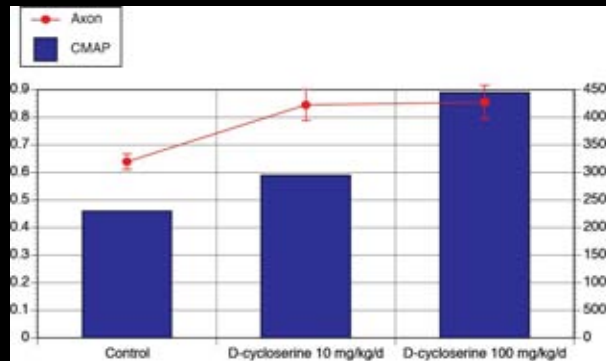


# Can we alter the rate of axonal elongation? - PNS

Unbiased screen for compounds that increased outgrowth in SC explants and DRG neurons



D-cycloserine  
A weak NMDA agonist



# Can we alter the rate of axonal elongation? - CNS

Phosphatase and tensin homologue (PTEN)

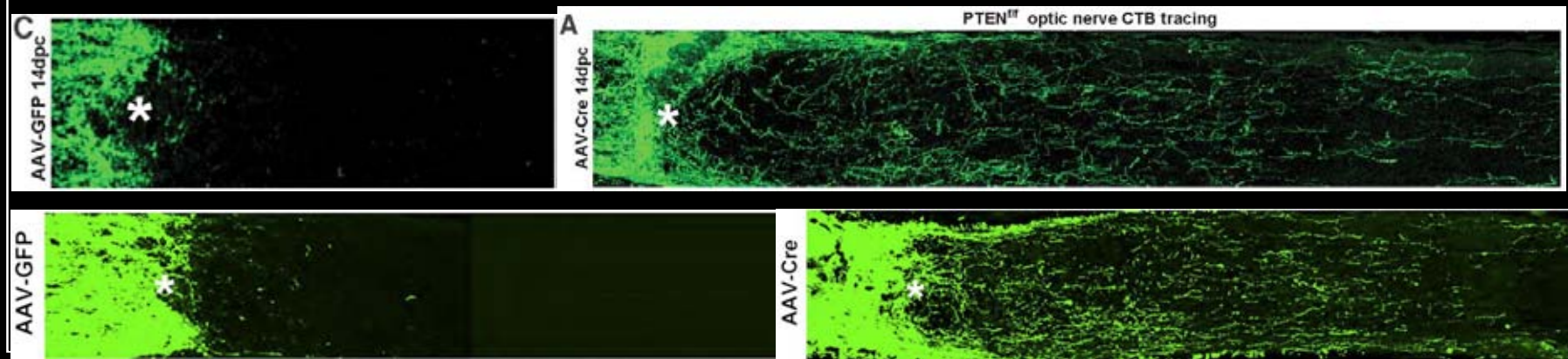
Mammalian target of rapamycin (mTOR)

Suppressor of cytokine signaling 3 (SOCS3)

Janus kinase/signal transducers and activators of transcription (JAK/STAT)

Optic nerve model of CNS regeneration:

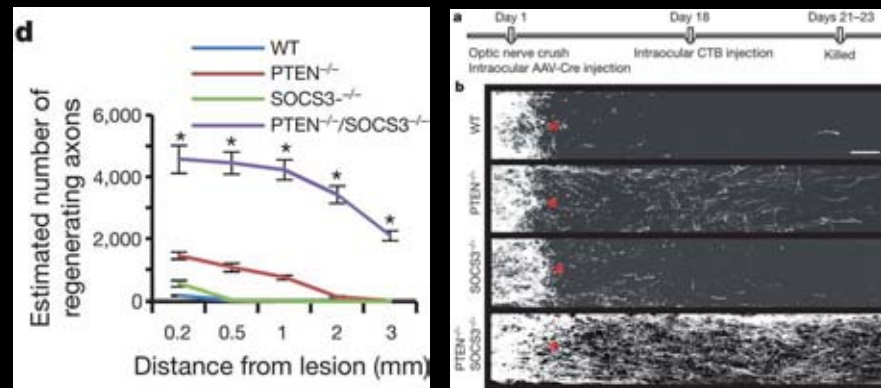
Adult retinal ganglion cells (RGCs)



# Can we alter the rate of axonal elongation? – CNS vs PNS

## Combined PTEN/SOCS3 deletion

Concurrent activation of mTOR and JAK/STAT pathways in neurons



(Sun et al 2011)

But no effect on PNS regeneration!

Intrinsic mechanisms of CNS vs PNS regeneration are likely to be different

Nevertheless, local inhibition of PTEN at site of regeneration accelerates PNS regeneration (Christie et al 2010)

# Gaps in our knowledge and future directions – ongoing studies

Chronic denervation in the:

Pathway: Schwann cells

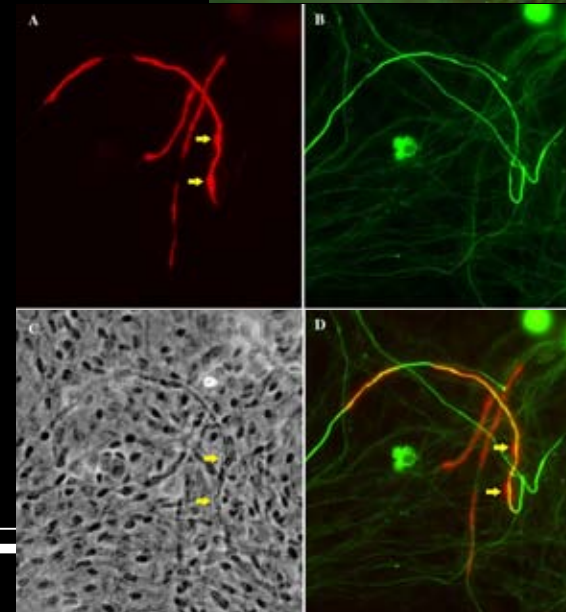
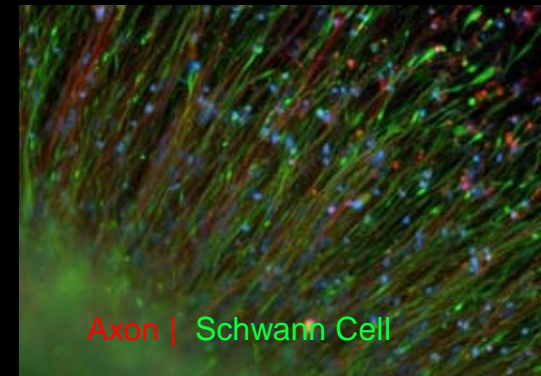
Target: muscle, skin

Can we reactivate atrophied SCs or prevent atrophy?

Role of neuregulin-1 type III

Do we need to replace atrophied SCs? Can we do it?

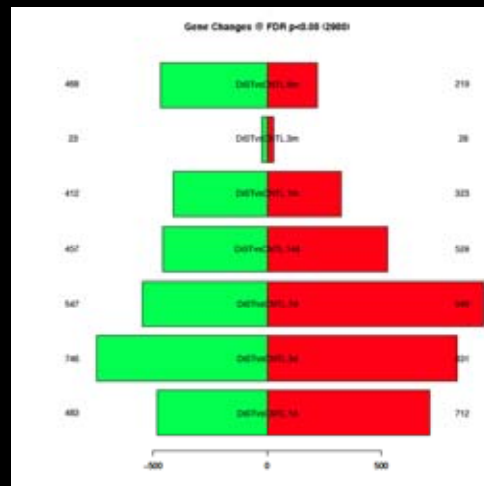
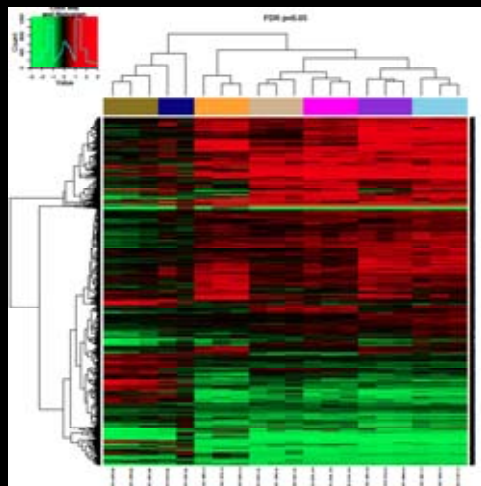
Human ESC or iPS generated Neural Crest Stem Cells as a source



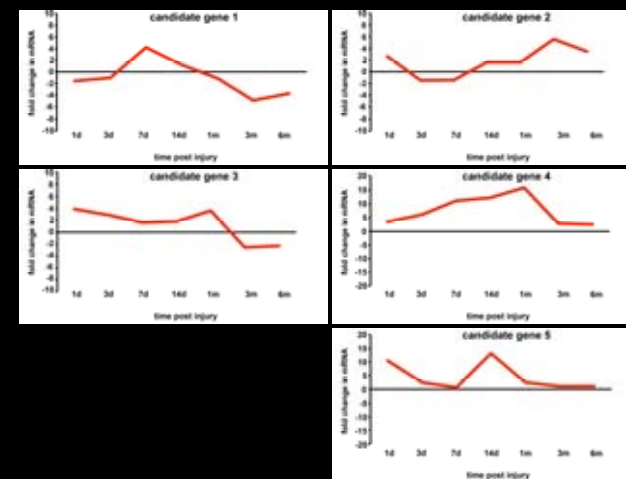


# Chronic denervation in the SCs

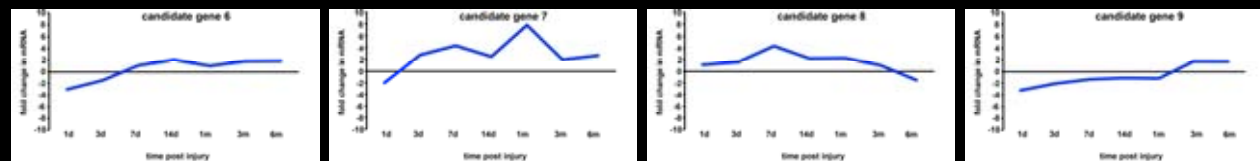
Rat sciatic nerve denervation 1d – 6 mo



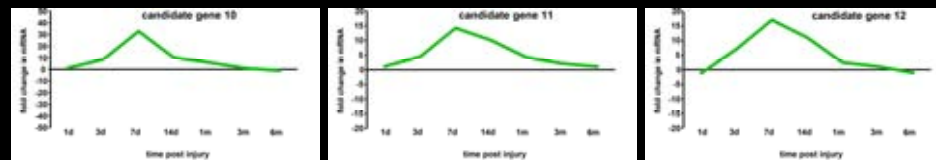
Transcription Factors



Apoptosis Related

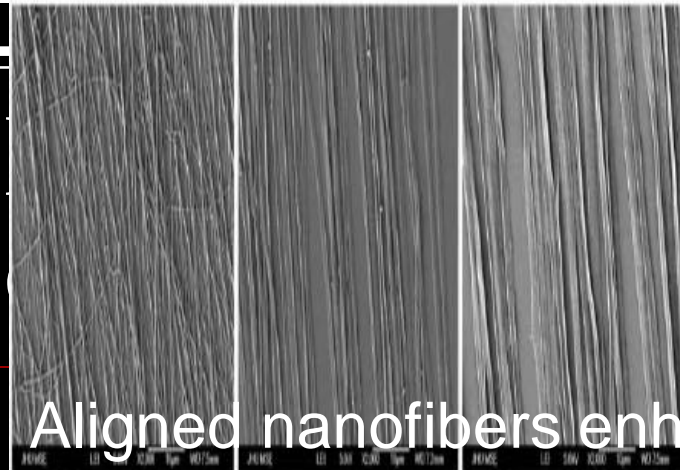


Cell-cycle proteins



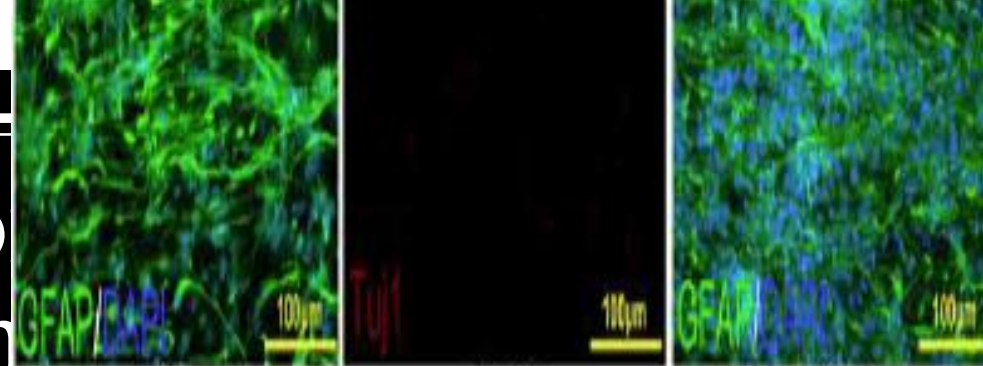
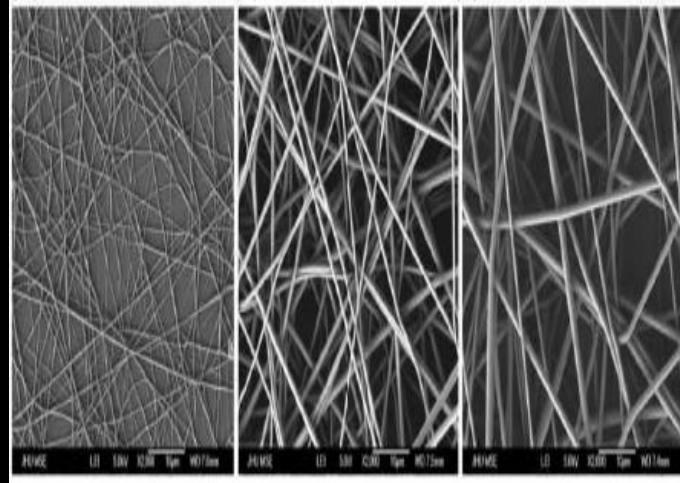
(Wright et al Unpublished)

(A) 160A:  $153 \pm 53$  nm (B) 600A:  $603 \pm 102$  nm (C) 1600A:  $1600 \pm 272$  nm

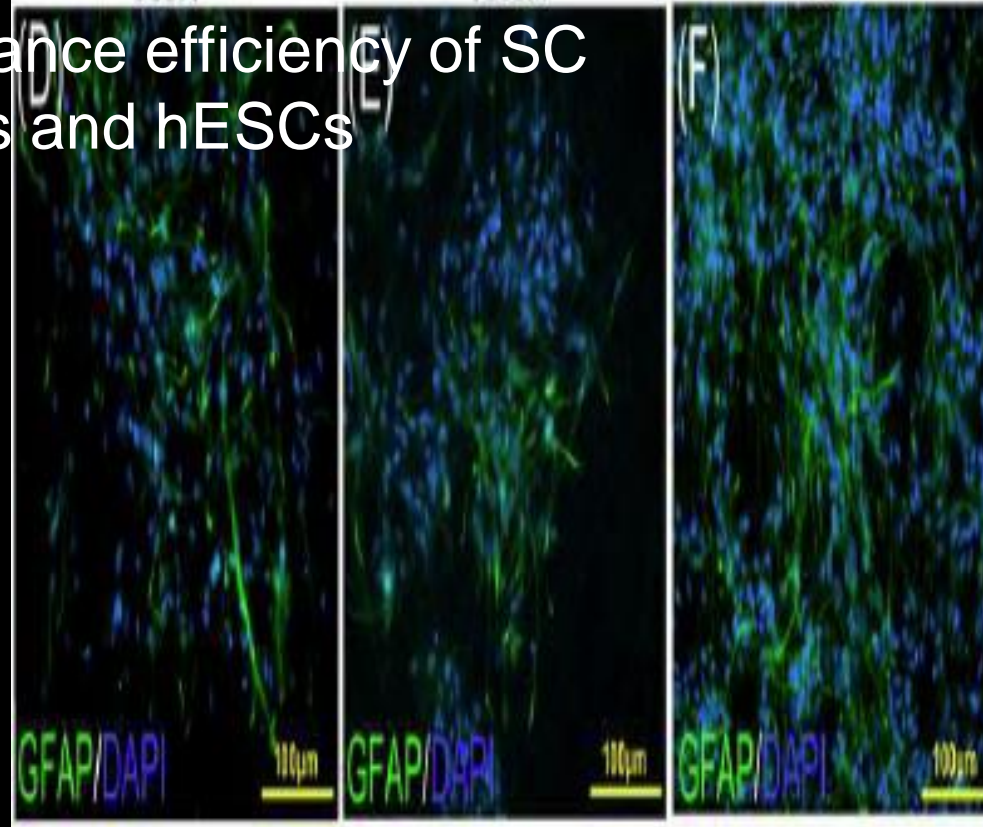


Aligned nanofibers enhance efficiency of SCs and hESCs

(D) 600R:  $160 \pm 46$  nm (E) 600R:  $546 \pm 87$  nm (F) 1600R:  $1570 \pm 448$  nm



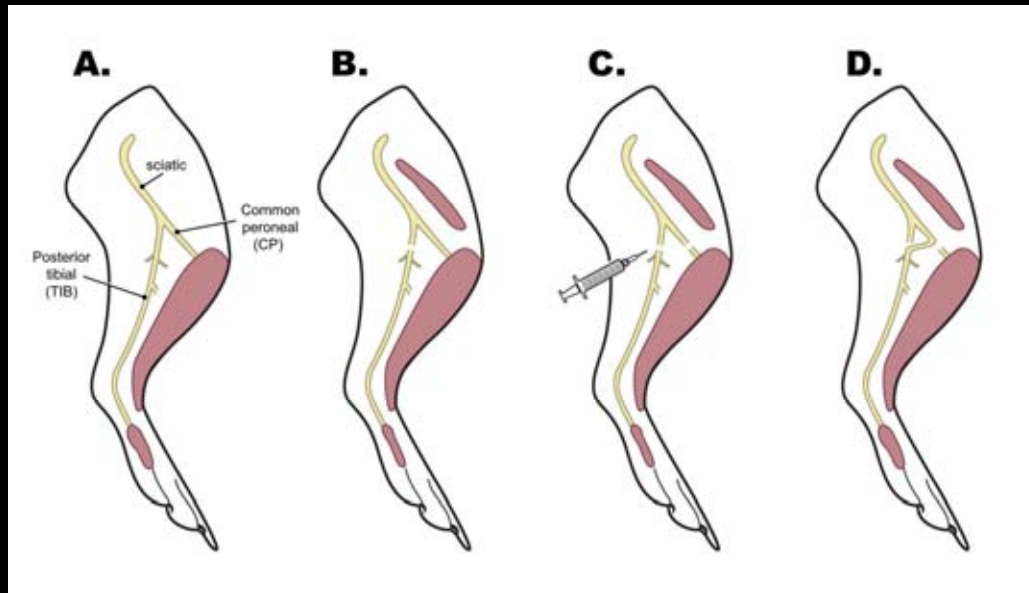
600R 1600R TCPS



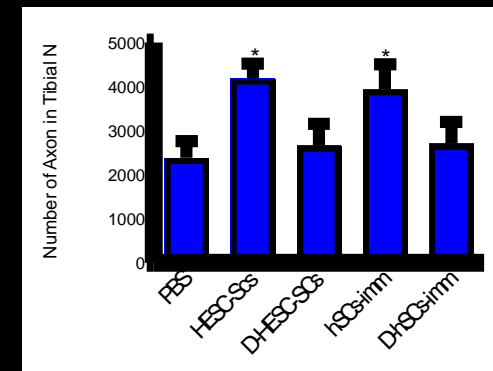
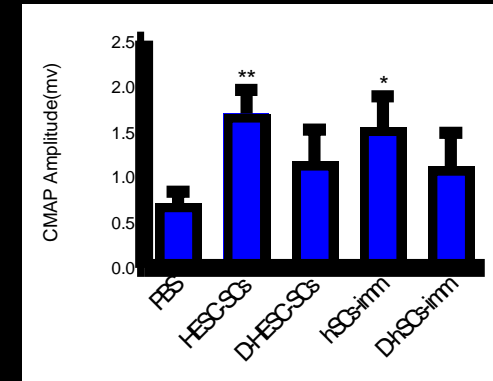
(Ren et al In Press)

# Replacement of chronically denervated Schwann cells

- hESC-derived Schwann cells enhance regeneration in chronically denervated nerves

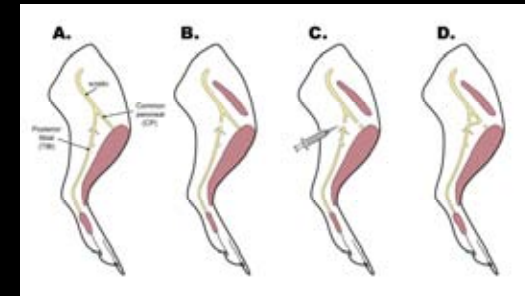
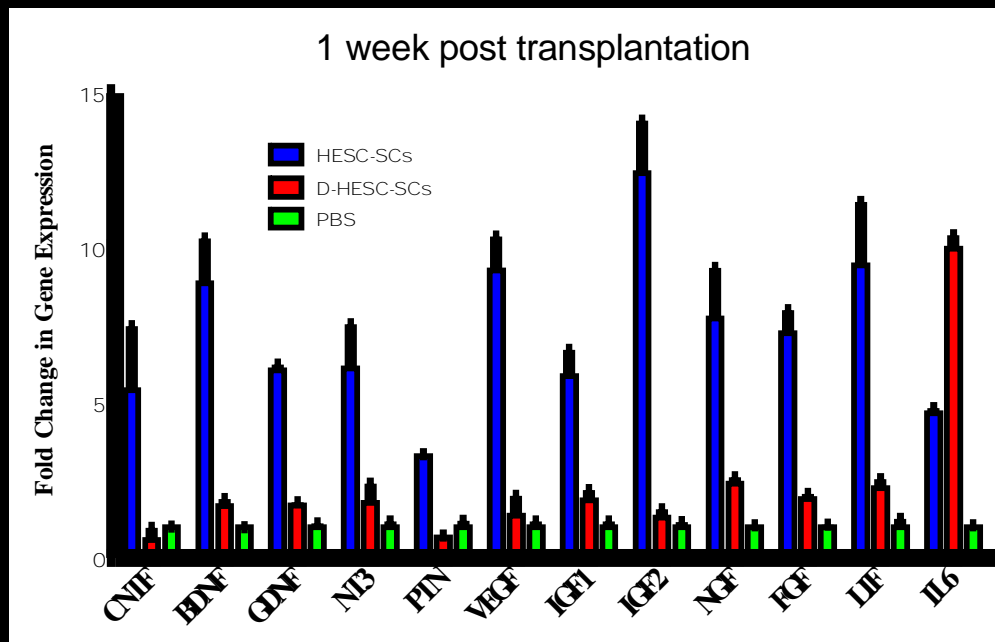


- But they do not survive



# Replacement of chronically denervated Schwann cells

- hESC-derived Schwann cells enhance regeneration in chronically denervated nerves
- Increased secretion of NTFs

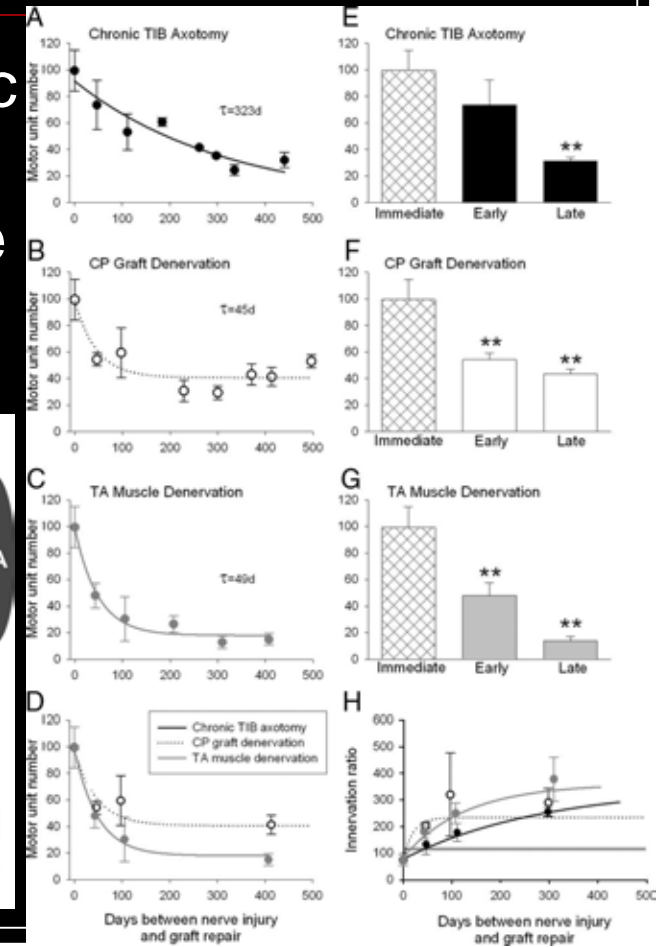
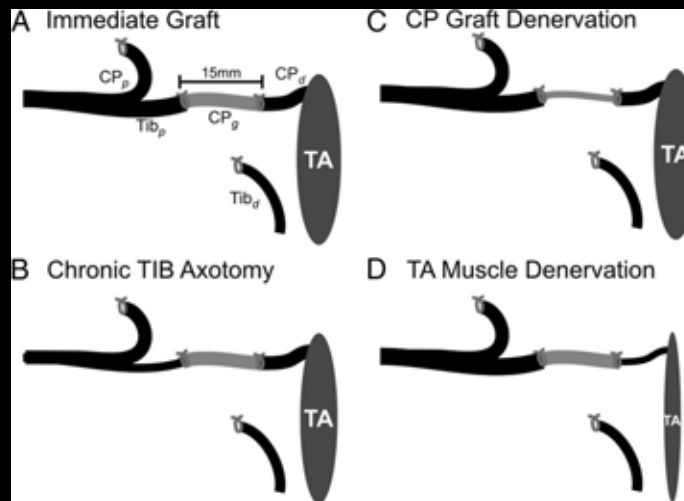


# Chronically denervated target: muscle

Even if we solve the issue of chronic denervation in Schwann cells, we still need to focus on changes at the NMJ and muscle

100-500 days of denervation

(Gordon et al J Neurosci 2011)



# Challenges in translating to humans

Do we have the appropriate biomarkers, tools to assess regeneration?

Nerve regeneration in humans is slow!!!

Issue of distance and rate of nerve regeneration in humans

We need validated biomarkers that can predict successful outcome before full recovery takes place

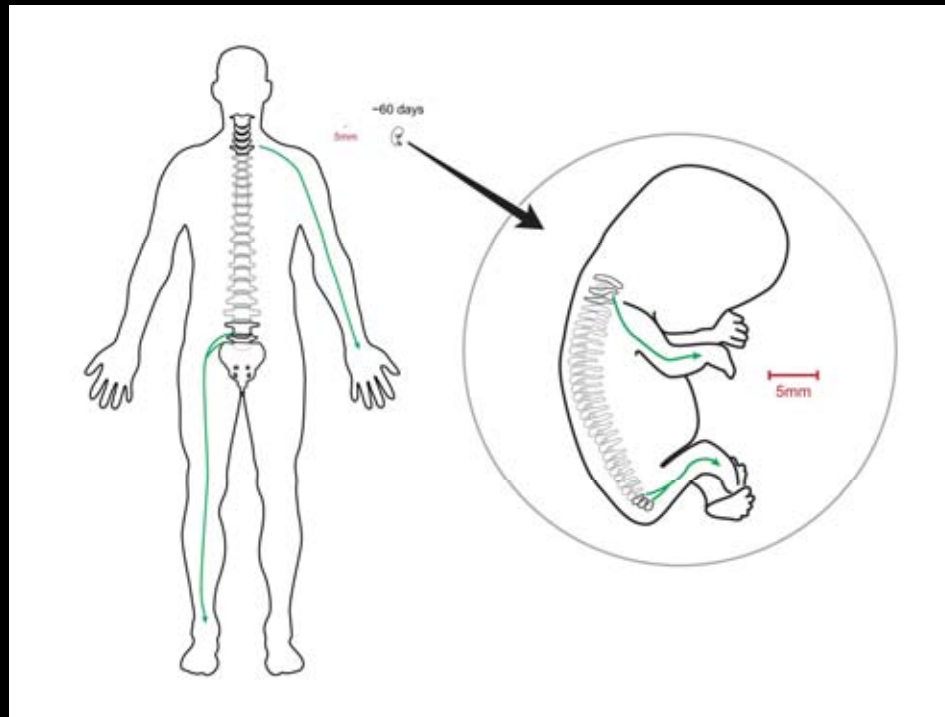
Novel human experimental assays

Imaging?



# Challenges in translating to humans

Regeneration is not same as development  
We need to make it faster and better



# Challenges in translating to humans

Do we have the appropriate biomarkers, tools to assess regeneration?

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# Biomarkers and tools to assess nerve regeneration

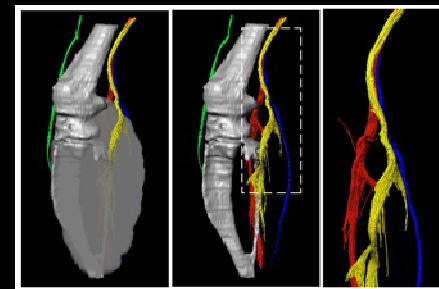
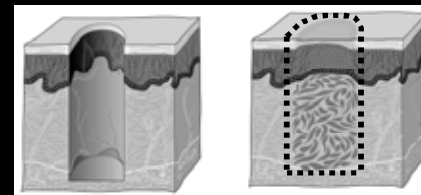
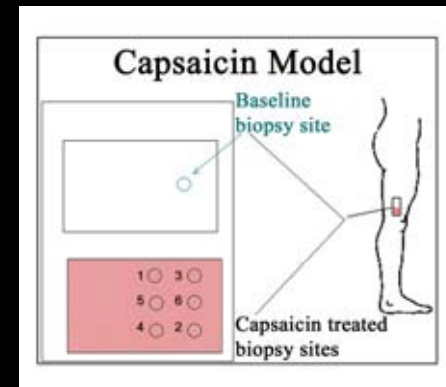
Clinical trials in peripheral nerve injuries are costly:

Nerve regeneration in humans is slow

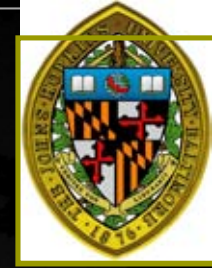
Novel models of assessing nerve regeneration in humans

Novel imaging techniques?

Diffusion Tensor Imaging



# Thanks



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- Ruifa Mi
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# References

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## § BOOKS

- § Brushart TM. Nerve Repair. Oxford: Oxford; 2011.
- § Zochodne DW. Neurobiology of peripheral nerve regeneration. Cambridge: Cambridge; 2008.

## § Reviews

- § Höke A. Mechanisms of Disease: what factors limit the success of peripheral nerve regeneration in humans? Nat Clin Pract Neurol 2006; 2: 448-454.
- § Experimental Neurology Special Issue: "Regeneration in the Peripheral Nervous System" Volume 223, Issue 1, pp. 1-250 (May 2010)
- § Höke, A. "A (heat) shock to the system promotes peripheral nerve regeneration" J Clin Invest. 2011 Vol: 121(11), pp:4231-4